21. Begun, D. R. Knuckle-walking ancestors. Science 259, 294 (1993).
22. Andrews, P. J. \& Martin, L. B. Cladistic relationships of extant and fossil hominoids. J. Hum. Evol. 16, 101-118 (1987).
23. Begun, D. R. Miocene fossil hominids and the chimp-human clade. Science 257, 1929-1933 (1992).
24. Ruvolo, M. Molecular phylogeny of the hominoids: inferences from multiple independent DNA sequence data sets. Mol. Biol. Evol. 14, 248-265 (1997).
25. Dainton, M. \& Macho, G. A. Did knuckle walking evolve twice? J. Hum. Evol. 36, 171-194 (1999).
26. Hunt, K. D. The postural feeding hypothesis: an ecological model for the origin of bipedalism. S. Afr. J. Sci. 9, 77-90 (1996).
27. Hewes, G. W. Food transport and the origin of hominid bipedalism. Am. Anthropol. 63, 687-710 (1961).
28. Lovejoy, C. O. The origin of man. Science 211, 341-350 (1981).
29. Napier, J. R. \& Davis, P. The forelimb skeleton and associated remains of Proconsul africanus. Foss. Mamm. Afr. 16, 1-70 (1959).
30. Grine, F. E. \& Susman, R. L. Radius of Paranthropus robustus from Member 1, Swartkrans Formation, South Africa. Am. J. Phys. Anthropol. 84, 229-248 (1991).

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# Predictive accuracy of population viability analysis in conservation biology 

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Population viability analysis (PVA) is widely applied in conservation biology to predict extinction risks for threatened species and to compare alternative options for their mangement ${ }^{1-4}$. It can also be used as a basis for listing species as endangered under World Conservation Union criteria ${ }^{5}$. However, there is considerable scepticism regarding the predictive accuracy of PVA, mainly because of a lack of validation in real systems ${ }^{2,6-8}$. Here we conducted a retrospective test of PVA based on 21 long-term ecological studies-the first comprehensive and replicated evaluation of the predictive powers of PVA. Parameters were
estimated from the first half of each data set and the second half was used to evaluate the performance of the model. Contrary to recent criticisms, we found that PVA predictions were surprisingly accurate. The risk of population decline closely matched observed outcomes, there was no significant bias, and population size projections did not differ significantly from reality. Furthermore, the predictions of the five PVA software packages were highly concordant. We conclude that PVA is a valid and sufficiently accurate tool for categorizing and managing endangered species.
PVA is a way to predict the probability of population (or species) extinction, by inputting actual life-history information and projecting it forward using stochastic computer simulation ${ }^{1-4}$. PVA is commonly used in a comparative way, to evaluate the effectiveness of different management options; because of this it has been argued that PVA predictions do not need to be precise ${ }^{2,9,10}$. However, there is no clear dichotomy between relative and absolute predictions-as conservation actions entail costs, management decisions are based not only on whether the proposed strategy is sufficient to achieve recovery, but also on whether the likely benefit will justify the expenditure. These considerations require PVA predictions to be quantitatively reliable. Uncertainties surrounding its predictive reliability have led to conclusions drawn from PVA being rejected in the law courts ${ }^{11}$. It is therefore essential that the predictions of PVA be compared and tested ${ }^{12-14}$. Here we assess the predictive accuracy of PVA, and determine whether different generic PVA computer packages differ in their predictive capabilities.

Historical data have been used to test and improve the predictions of complex climatic ${ }^{15}$, economic ${ }^{16}$, geological ${ }^{17}$ and ecological ${ }^{18}$ models. As PVA models have important stochastic components, the conclusions of past studies based on a single test ${ }^{13,19}$ lack power, and are prone to case-specific peculiarities ${ }^{20}$. Consequently, a valid test of PVA predictions must incorporate a large number of data sets to obtain representative assessments. We conducted retrospective analyses on 21 wildlife populations- 8 avian, 11 mammalian (representing 9 species), 1 reptilian and 1 piscine.

The 21 data sets used were the only long-term studies we identified that presented data of sufficient duration and quality to be suitable for retrospective testing (see Methods). Five of the most commonly applied 'generic' PVA packages (GAPPS, INMAT, RAMAS Metapop, RAMAS Stage and VORTEX) were used. These packages are all suitable for generic, single-population risk assessments. They offer the most realistic prospects for improving PVA as they are subject to wide scrutiny, are repeatedly used and are frequently revised and updated. All have been used in the management and conservation of endangered species. The key features and differences between the five PVA packages are given in the Supplementary Information.

To avoid circularity, the total data available for each population was split. The first half was used to develop and parameterize the models; the latter half was reserved for testing the accuracy of the PVA predictions. To ensure that the two time periods were kept completely separate, no information from the second half of the population history was used in formulating or parameterizing the

|  | Expected number | GAPPS* | INMAT | R META | R STAGE | VORTEX |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| No. $N($ actual $)<N(Q 90 \%)$ | $\begin{gathered} 18.9 \\ \text { G-test } P \end{gathered}$ | $\begin{aligned} & 17 \\ & 0.48 \end{aligned}$ | $\begin{aligned} & 16 \\ & 0.07 \end{aligned}$ | $\begin{aligned} & 19 \\ & 0.94 \end{aligned}$ | $\begin{aligned} & 17 \\ & 0.21 \end{aligned}$ | $\begin{aligned} & 17 \\ & 0.21 \end{aligned}$ |
| No. $N($ actual $)<N(Q 50 \%)$ | $\begin{gathered} 10.5 \\ G \text {-test } P \end{gathered}$ | $\begin{aligned} & 11 \\ & 0.65 \end{aligned}$ | $\begin{aligned} & 10 \\ & 0.83 \end{aligned}$ | $\begin{aligned} & 8 \\ & 0.27 \end{aligned}$ | $\begin{gathered} 11 \\ 0.83 \end{gathered}$ | $\begin{aligned} & 8 \\ & 0.27 \end{aligned}$ |
| No. $N$ (actual) < N(Q10\%) | $\begin{gathered} 2.1 \\ \text { G-test } P \end{gathered}$ | $\begin{aligned} & 3 \\ & 0.48 \end{aligned}$ | $\begin{aligned} & 1 \\ & 0.38 \end{aligned}$ | $\begin{aligned} & 3 \\ & 0.54 \end{aligned}$ | $\begin{aligned} & 4 \\ & 0.21 \end{aligned}$ | $\begin{aligned} & 3 \\ & 0.54 \end{aligned}$ |

 packages GAPPS, INMAT, RAMAS Metapop, RAMAS Stage and VORTEX. The sample is based on 21 retrospective studies. None of the PVA packages' predictions differed significantly from the expected outcome based on goodness of fit ( $G$ ) tests.
*GAPPS crashes at very large population sizes because of memory limitations, so the fish was not modelled with this package (expected numbers adjusted to 18, 10 and 2).



Figure 1 Plot of the PVA-predicted probability of population decline (quasi-extinction risk) versus the actual proportion of the 21 real populations that decline below the corresponding threshold size. These threshold sizes represent different percentage declines in different populations, but are always associated with the same level of risk. For example, half (10.5) of the 21 historical populations should have actually declined below the size assigned a 50\% probability by the PVA. For each of the five PVA software packages, a perfect fit with reality lies on the $45^{\circ}$ line.

PVA models. All predictive tests were done within species and then pooled across taxa. The accuracy of PVA was assessed by comparing the predicted quasi-extinction risk (the likelihood that the population will decline below a given size in a specified time frame ${ }^{21}$ ) and population size projections with reality (see Methods).

The results showed a surprisingly close relationship between the PVA predictions and the historical behaviours of the 21 real populations. The quasi-extinction probabilities predicted by the PVAs were not significantly different from the proportion of actual populations that declined below a given threshold (Fig. 1). The risks of decline predicted by the PVAs did not deviate significantly from reality (Table 1), although there was a trend towards slightly pessimistic predictions. Furthermore, there were no overall differences between PVA software packages in their predictions of quasiextinction risks at $90 \%, 50 \%$ and $10 \%$ risk levels (for example, at the $50 \%$ quasi-extinction level, Kruskall-Wallis rank test, $H=1.01$, d.f. $=4, P=0.907$ ).

Actual population sizes consistently fell within the bounds predicted by the stochastic simulations. The projected mean final population sizes were not significantly different from the actual final population sizes (for any of the PVA packages), on the basis of a distribution test combining all 21 studies. Likewise, predicted and actual population growth rates $(r)$ did not differ significantly. There was no consistent over- or underestimation of future population numbers (Fig. 2). A sign test on the predictive bias was not significant for any package, nor when pooled across all packages. There was also no difference in bias across the five PVA packages $(G[$ Heterogeneity $]=0.957$, d.f. $=4, P=0.916)$.

The predictions of the five PVA packages were significantly correlated across the 21 retrospective studies, on the basis of both quasi-extinction probabilities and the fit of projections to future population sizes (Table 2). There were some significant differences


Figure 2 Signed predictive bias in projected population size (compared with actual population numbers), taken across 21 populations for five PVA packages. There is no significant positive or negative bias.
between particular packages for individual species (on the basis of Kolmogorov-Smirnov confidence interval tests ${ }^{22}$ on quasi-extinction probabilities). However, there was no overall consistency in these differences, and no package gave demonstratively better or worse predictions than any other. RAMAS Metapop and VORTEX gave the best least-squares model fit to actual population size (taken across all 21 species; see Table 3 of Supplementary Information), although a Kruskall-Wallis rank test showed no significant difference among the PVA packages $(H=7.9$, d.f. $=4, P=0.097)$.

The short-term predictions of the PVA models were relatively accurate, with a good overall correspondence between simulated and observed outcomes, on the basis of several criteria. The PVA predictions were not overly optimistic, as has been assumed by some ${ }^{10}$ (in fact, the general trend was for the PVA models to be slightly pessimistic). Following the precautionary principle, it is preferable to produce circumspect predictions of risk when managing endangered species ${ }^{23}$. These findings contradict the widespread view that stochastic population models are poor predictors of a population's future fate. Nevertheless, if the means and/or variances of vital rates were to change substantially in the future, the absolute accuracy of PVA predictions would be questionable. Furthermore, these results may not apply to plants (which were not included in this study), nor to cases where too few life-history or population data are available to estimate model structure and input parameters reliably ${ }^{13}$.

In some cases, there were significant absolute differences between the predictions of different software packages, due to differences in model structure. For example, a ceiling carrying capacity had to be implemented for the cycling Soay and Boreray sheep populations in INMAT, even though this was known to be inappropriate for these populations ${ }^{24}$. In these cases, the other packages that allowed overcompensatory density-dependent survival to be modelled gave more realistic predictions than INMAT. For a given species, the best packages are likely to be those that most realistically model its life-history. Unfortunately, it is not always possible to discern the type of model that fits a particular case a priori. Despite the best efforts of PVA modellers, it is possible that some events (such as catastrophes) may be overlooked, making it impossible to guarantee

Table 2 Correlations between the predictions of five PVA packages

|  | Probability of decline* |  |  |  | Model fit to actual $\mathrm{N} \dagger$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | INMAT | R META | R STAGE | VORTEX | INMAT | R META | R STAGE | VORTEX |
| GAPPS | 0.83 | 0.78 | 0.86 | 0.86 | 0.66 | 0.94 | 0.88 | 0.90 |
| INMAT |  | 0.63 | 0.87 | 0.76 |  | 0.70 | 0.82 | 0.78 |
| R META |  |  | 0.83 | 0.87 |  |  | 0.94 | 0.89 |
| R STAGE |  |  |  | 0.88 |  |  |  | 0.89 |

*Probability of the maximum observed historici....................................... population decline.
$\dagger$ Scaled fit of model projections to actual population size $(N)$.
All correlations are highly significant $(P<0.01)$.
that incorrect predictions will not be made. It is therefore important to explore alternative model structures and conduct sensitivity analyses, particularly when data are scarce ${ }^{2,3,14}$. Moreover, information collected from past monitoring should be used to test and refine PVA models, forming a continual feedback process of development and improvement ${ }^{25}$. PVA is the best tool we have for estimating extinction risk, and the alternatives are subjective, less rigorous, and likely to provide poorer predictions ${ }^{26}$.

PVA predictions are surprisingly accurate, given adequate data, and should be useful in the conservation contexts in which they are currently applied. There are also high correlations between the predictions of different PVA packages. These results provide strong empirical justification for the use of PVA for categorizing the vulnerability of endangered animal species and evaluating options for their recovery. Furthermore, they validate PVA as a useful research tool for addressing unresolved issues in conservation biology.

## Methods

## Data sets

The protocol for choosing examples was independent of their structure, detail and outcome. The criteria used were: (1) a minimum duration of 10 years; (2) data of sufficient quality and detail to build PVA models; (3) a small population size (very large populations are generally not of conservation concern); (4) data concerning endangered species or isolated populations. Thus, the filter was based on the priorities of our colleagues in collecting ecological data. The 21 populations that we were able to use (see Supplementary Information) covered a range of taxa (birds, mammals, reptiles and fish), tropic levels (omnivores, herbivores and carnivores), environmental conditions (low to high environmental variability) and insular and mainland populations, encompassed several different modes of population regulation (exponential growth or decline, a population ceiling, and density dependence), and spanned a range of geographical and climatic zones (Africa, Europe, North America and Oceania). Nevertheless, not all taxa are completely or proportionally represented in our sample, and the focus is on extant species.

## PVA model structure

The structure of each PVA model depended on the biology of the given species and the built-in features of each software package (see Supplementary Information). To ensure that the models were as realistic as possible, all relevant aspects of the population's ecology were included (within the capabilities of each PVA package). The exact structure of the PVA models built using different packages often differed for a given species, depending on the available features, limitations and assumptions associated with each program ${ }^{27}$. This made it possible to test whether some packages were better predictors than others because of their different features.

## Parameter estimation

We used the best estimates for each parameter, given the available demographic, environmental and population data. Where the estimated population parameters were given directly in the literature, we checked them independently when possible. In most cases the compilation and analysis of data were done in collaboration with the people who worked on the taxon in question. The protocols used to estimate the PVA model parameters are presented in the Supplementary Information.

## PVA runs

Five hundred stochastic simulation runs were performed in each case to ensure statistical reliability.

## Testing PVA predictions

The predictive accuracy of PVA was assessed as follows. (1) Quasi-extinction probabilities were used as a surrogate for absolute extinction risk, as only one study population went extinct. If the PVA predictions are correct, the proportion of historical populations that declined below a given threshold should equate with the proportion of simulation trajectories that declined below the same threshold. To test this, the number of actual populations (out of 21) that declined below the threshold population sizes corresponding to predicted quasi-extinction probabilities of $0 \%, 10 \%, 20 \%, \ldots, 100 \%$ were recorded. (2) Mean population size predictions and growth rates were compared with reality using a combined test based on the sum of standard deviates ${ }^{28}$. The objective was to test for equality over multiple populations, and combine the results of individual paired tests to increase statistical power. A log-transformation was used to normalize population size data. (3) Signed predictive bias (mean error): models with good predictive properties will produce a bias close to zero. Bias was calculated as $\Sigma$ (Predicted size - Actual size)/ number of years $(N)$. This was converted to a normal deviate $W=\sqrt{ } N \times$ bias $/ s$, where $s$ is the standard deviation of bias values. (4) The 'fit' of predicted versus actual population size over time was used to assess projections of future population numbers, calculated as the sum of squared deviations between observed and actual population sizes
( $\Sigma[\text { Predicted }- \text { Actual }]^{2} /$ Actual $)$ for each simulation replicate, averaged over 500 repli-
cates. (5) Pearson's product moment correlations were used to determine concordance amongst the predictions of different PVA packages for quasi-extinction risk and population size projections.

## Statistical assumptions

This study represents a general 'meta-analysis' problem ${ }^{29}$, the units of analysis being independent studies rather than individual subjects. The statistical population was composed of all studied biological populations with sufficient data, and was sampled completely. The underlying assumption of all our statistical tests on the combined results was that the 'errors' were independent among the 21 cases $^{22}$. As the case studies had little overlap in terms of place, time-period or researcher, this is a valid assumption. The quasiextinction analysis was based on a simple goodness of fit test, carrying no assumption about the distributions of individual cases. For tests comparing different PVA packages, we verified that the assumptions of parametric tests ${ }^{13,14}$ were met.

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1. Boyce, M. S. Population viability analysis. Annu. Rev. Ecol. Syst. 23, 481-506 (1992).
2. Beissinger, S. R. \& Westphal, M. I. On the use of demographic models of population viability in endangered species management. J. Wildl. Mgmt. 62, 821-841 (1998).
3. Groom, M. J. \& Pascual, M. A. in Conservation Biology (eds Fiedler, P. L. \& Kareiva, P. M.) 4-27 (Chapman \& Hall, New York, 1998).
4. Norton, T. W. (ed.) Special issue: applications of population viability analysis to biodiversity conservation. Biol. Conserv. 73, 91-176 (1995).
5. International Union for the Conservation of Nature. IUCN Red List Categories (Gland, Switzerland, 1994).
6. Caughley, G. Directions in conservation biology. J. Anim. Ecol. 63, 215-244 (1994).
7. Taylor, B. L. The reliability of using population viability analysis for risk classification of species. Conserv. Biol. 9, 551-558 (1995).
8. Ludwig, D. Is it meaningful to estimate a probability of extinction? Ecology 80, 298-310 (1999).
9. Possingham, H. P., Lindenmayer, D. B. \& Norton, T. W. A framework for the improvement of threatened species based on population viability analysis (PVA). Pac. Conserv. Biol. 1, 38-45 (1993).
10. Ralls, K. \& Taylor, B. L. in The Ecological Basis of Conservation: Heterogeneity, Ecosystems, and Biodiversity (eds Pickett, S. B. A., Ostfeld, R. S., Shachak, M. \& Likens, G. E.) 228-235 (Chapman \& Hall, New York, 1997).
11. Talbot, J. Land and Environment Court of New South Wales: Record of Hearing \#10151 of 1994 (Court Report, Sydney, 1994).
12. Soulé, M. E. \& Kohm, K. A. Research Priorities for Conservation Biology (Island, Washington DC, 1989).
13. Brook, B. W., Lim, L., Harden, R. \& Frankham, R. Does population viability analysis software predict the behaviour of real populations? A retrospective study on the Lord Howe Island woodhen Tricholimnas sylvestris (Sclater). Biol. Conserv. 82, 119-128 (1997).
14. Brook, B. W., Cannon, J. R., Lacy, R. C., Mirande, C. \& Frankham, R. A comparison of the population viability analysis packages GAPPS, INMAT, RAMAS and VORTEX for the Whooping crane (Grus americana). Anim. Conserv. 2, 23-31 (1999).
15. Wigley, T. M. L. Climate change: a successful prediction? Nature 376, 463-464 (1995).
16. Ramanathan, R. Introductory Econometrics with Applications 2nd edn (Dryden, Fort Worth, 1995).
17. Oreskes, N., Shrader-Frenchette, K. \& Belitz, K. Verification, validation, and the confirmation of numerical models in the earth sciences. Science 263, 641-646 (1994).
18. Wahlberg, N., Moilanen, A. \& Hanski, I. Predicting the occurrence of endangered species in fragmented landscapes. Science 273, 1536-1538 (1996).
19. Huggard, D. J. A linear programming model of herbivore foraging: imprecise, yet successful? Oecologia 100, 470-474 (1994).
20. Belovsky, G. E. How good must models and data be in ecology? Oecologia 100, 475-480 (1994).
21. Ginzburg, L. R., Slobodkin, L. B., Johnson, K. \& Bindman, A. G. Quasiextinction probabilities as a measure of impact on population growth. Risk Anal. 2, 171-182 (1982).
22. Sokal, R. R. \& Rohlf, F. J. Biometry 4th edn (Freeman, New York, 1995).
23. Akçakaya, H. R., Burgman, M. A. \& Ginzburg, L. R. Applied Population Ecology 2nd edn (Sinauer, Sunderland, Massachusetts, 1999).
24. Grenfell, B. T., Price, O. F., Albon, S. D. \& Clutton-Brock, T. H. Overcompensation and population cycles in an ungulate. Nature 355, 823-826 (1992).
25. Burgman, M. A., Ferson, S. \& Akçakaya, H. R. Risk Assessment in Conservation Biology (Chapman \& Hall, London, 1993).
26. Zeckhauser, R. J. \& Viscusi, W. K. Risk within reason. Science 248, 559-564 (1990).
27. Mills, L. S. et al. Factors leading to different viability predictions for a grizzly bear data set. Conserv. Biol. 10, 863-873 (1996).
28. Reynolds, M. R., Burkhart, H. E. \& Daniels, R. F. Procedures for statistical validation of stochastic simulation models. Forest Sci. 27, 349-364 (1981).
29. Arnqvist, G. \& Wooster, D. Meta-analysis: synthesizing research findings in ecology and evolution. Trends Ecol. Evol. 10, 236-240 (1995).

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## Supplementary Information

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## Features of the PVA packages

Table 1 summarises the capabilities and limitations of the five generic PVA packages used.

## Protocols used to estimate PVA input parameters

Demographic structure: Age was used as the standard demographic partition. Number of classes modelled depended on whether the vital rates (fecundity and survival) were age dependent or constant with age. The initial population was set up in a stable age distribution ${ }^{1}$ unless the actual distribution was known.

Survival rates: Proportion of individuals that survived from age $x$ to age $x+1$, estimated by weighted averages. Where sufficient data existed, sex-specific rates were used in the individual-based packages. The maximum age represented an absolute ceiling on survival.
Reproductive rates: Fecundity was estimated as the average number of offspring per adult (matrixbased models), or the number of male and female offspring per female (individual-based models). Litter size was specified as a Poisson distribution. For pre-breeding census data, fecundity incorporated juvenile survival. For post-breeding census data, adult survival prior to breeding was included ${ }^{1}$. Age of parturition and breeding systems (monogamous, polygamous etc.) were procured from the literature.
Density dependence ( $D D$ ): To determine the form of DD operating (if any), we relied on the biological and statistical evidence given in the literature, time-series tests ${ }^{2}$, and regressions of the vital rates on population size. A non-linear least-squares regression algorithm was used to find the best parameters for DD models. A population ceiling was used in cases where evidence for DD effects on the vital rates was not significant, but there was a definite population size limit governed by the amount of available habitat and resources or the number of territories.
Environmental variation (EV): Annual variability was estimated as the standard deviation of the average vital rates, after removing the expected variance produced by chance demographic sampling ${ }^{3}$. This avoids duplicating the effects of demographic stochasticity, which is generated internally by the PVA packages. When functional DD was modelled, EV $=\sqrt{ }\left(\sum\left(y_{i}-f\left(\mathrm{x}_{\mathrm{i}}\right)\right)^{2} /(n-1)\right)$, where $\mathrm{y}_{\mathrm{i}}$ was the actual value for a given value of $\mathrm{x}_{\mathrm{i}}, f\left(\mathrm{x}_{\mathrm{i}}\right)$ was the DD function estimated for y from $\mathrm{x}_{\mathrm{i}}$, and $n$ was the number of observations. This separated deterministic effects from stochastic fluctuations.
Catastrophes: Extreme events that fell outside the typical regime of EV were simulated separately as catastrophes ${ }^{4}$ when a valid cause (e.g. cyclones) could be identified. Frequency = \# catastrophe years / \# census years. Impact = vital rate (survival and/or fecundity) in catastrophe year / average vital rate.
Inbreeding depression: In demographically isolated populations, where migration could not mitigate inbreeding effects, inbreeding depression is likely to be an important factor ${ }^{5,6}$. Quantitative data on the impact of inbreeding depression rarely existed, so we applied a value of 3.14 lethal equivalent alleles on
juvenile survival ${ }^{7}$. This is likely to be conservative, as it impacts on only one component of fitness, yet effects on all aspects of reproductive fitness are expected.

## Life-history data and parameter estimates

Tables 2 and 3 provide a summary of information on population data, life-history details and PVA model parameter estimates for the 21 populations examined in the retrospective evaluation.

## Fit of PVA models to each population

Table 4 shows the least-squares fit of each PVA package (ranked relative to the other packages) and the bias in projected population size, for each of the 21 populations. Although the PVA predictions of future population size were relatively accurate for the majority of species, in the case of the Isle Royale wolf population the fit was noticeably poor. The wolf population increased throughout the first half of it's monitored history, but later crashed due to an outbreak of canine parvovirus ${ }^{8}$. A projection based on the extrapolation of past trends did not anticipate this catastrophic event, and therefore did not predict such a sudden drastic drop in numbers. Clearly when qualitative changes occur in a species' dynamics, and these shifts are unforeseen by the modeller or biologist, the resulting PVA projections are likely to be inaccurate. However, once the possibility (of a rare outbreak) is known, it can be modelled as a catastrophe, which would avoid underestimating risk.

1. Caswell, H. Matrix Population Models: Construction, Analysis, and Interpretation (Sinauer, Sunderland, 1989).
2. Gaston, K. J. \& Lawton, J. H. A test of statistical techniques for detecting density dependence in sequential censuses of animal populations. Oecologia 74, 404-410 (1987).
3. Kendall, B. E. Estimating the magnitude of environmental stochasticity in survivorship data. Ecol. Applic. 8, 184193 (1998).
4. Lande, R. Risks of population extinction from demographic and environmental stochasticity, and random catastrophes. Am. Nat. 142, 911-927 (1993).
5. Frankham, R. \& Ralls, K. Inbreeding leads to extinction. Nature 392, 441-442 (1998).
6. Soulé, M. E. \& Mills, L. S. No need to isolate population genetics. Science 282, 1658-1659 (1998).
7. Ralls, K., Ballou, J. D. \& Templeton, A. Estimates of lethal equivalents and the cost of inbreeding in mammals. Conserv. Biol. 2, 185-193 (1988).
8. Peterson, R. O., Thomas, N. J., Thurber, J. M., Vucetich, J. A. \& Waite, T. A. Population limitation and the wolves of Isle Royale. J. Mammal. 79, 828-841 (1998).
9. Harris, R. B., Metzgar, L. H. \& Bevins, C. D. GAPPS: Generalized Animal Population Projection System (Univ. Montana, Missoula, 1986).
10. Mills, L. S. \& Smouse, P. E. Demographic consequences of inbreeding in remnant populations. Am. Nat. 144, 412-431 (1994).
11. Akçakaya, H. R. RAMAS ${ }^{\circledR}$ Metapop: Viability Analysis for Stage-structured Metapopulations (Applied Biomathematics, Setauket, 1997).
12. Ferson, S. RAMAS ${ }^{\circledR}$ Stage: Generalized Stage-based Modeling for Population Dynamics (Applied Biomathematics, Setauket, 1994).
13. Lacy, R. C. Vortex: a computer simulation model for population viability analysis. Wildl. Res. 20, 45-65 (1993).
14. Komdeur, J. Breeding of the Seychelles magpie robin Copsychus sechellarum and implications for its conservation. Ibis 138, 485-491 (1996).
15. Mirande, C. et al. Computer simulations of possible futures for two flocks of whooping cranes. Proc. 7th N. Amer. Crane Workshop 7, 181-200 (1997).
16. Møller, A. P. Population dynamics of a declining swallow Hirundo rustica population. J. Anim. Ecol. 58, 10511063 (1989).
17. Arcese, P., Smith, J. N. M., Hochachka, W. M., Rogers, C. M. \& Ludwig, D. Stability, regulation, and the determination of abundance in an insular song sparrow population. Ecology 73, 805-822 (1992).
18. Valle, C. A. Effective population size and demography of the rare flightless Galápagos cormorant. Ecol. Applic. 5, 601-617 (1995).
19. Loery, G. \& Nichols, J. D. Dynamics of a black-capped chickadee population, 1958-1983. Ecology 66, 11951203 (1985).
20. Brook, B. W., Lim, L., Harden, R. \& Frankham, R. How secure is the Lord Howe Island woodhen? A population viability analysis using VORTEX. Pac. Conserv. Biol. 3, 125-133 (1997).
21. Brook, B. W. \& Kikkawa, J. Examining threats faced by island birds: a PVA on the Capricorn silvereye using long-term data. J. Appl. Ecol. 35, 491-503 (1998).
22. Vucetich, J. A., Peterson, R. O. \& Waite, T. A. Effects of social structure and prey dynamics on extinction risk in gray wolves. Conserv. Biol. 11, 957-965 (1997).
23. Boyd, I. L. Population changes and the distribution of a herd of feral goats (Capra sp.) on Rhum, Inner Hebrides, 1960-78. J. Zool., Lond. 193, 287-304 (1981).
24. Lowe, V. P. W. Population dynamics of the red deer (Cervas elephus L.) on Rhum. J. Anim. Ecol. 38, 425-457 (1969).
25. Clutton-Brock, T. H., Guinness, F. E. \& Albon, S. D. Red Deer: The Behaviour and Ecology of Two Sexes (Univ. Chicago Press, 1982).
26. Hoogland, J. L. The Black-Tailed Prairie Dog: Social Life of a Burrowing Mammal (Univ. Chicago Press, 1995).
27. Burrows, R., Hofer, H. \& East, M. L. Demography, extinction and intervention in a small population: the case of the Serengeti wild dogs. Proc. R. Soc. Lond. B 256, 281-292 (1994).
28. Schwartz, O. A., Armitage, K. B. \& Van Vuren, D. A 32-year demography of yellow-bellied marmots. J. Zool., Lond. 246, 337-346 (1998).
29. McCullough, D. R. The George Reserve Deer Herd: Population Ecology of a K-selected Species (Univ. Michigan Press, Ann Arbor, 1979).
30. Clutton-Brock, T. H. et al. Population fluctuations, reproductive costs and life-history tactics in female Soay Sheep. J. Anim. Ecol. 65, 675-689 (1996).
31. Eberhardt, L. L., Blanchard, B. M. \& Knight, R. R. Population trend of the Yellowstone grizzly bear as estimated from reproductive and survival rates. Can. J. Zool. 72, 360-363 (1994).
32. McFadden, J. T., Alexander, G. R. \& Shetter, D. S. Numerical changes and population regulation in brook trout Salvelinus fontinalis. J. Fish. Res. 24, 1425-1459 (1967).
33. Tinkle, D. W., Dunham, A. E. \& Congdon, J. D. Life history and demographic variation in the lizard Sceloporus graciosus: a long-term study. Ecology 74, 2413-2429 (1993).

Table 1 List of modelling features available for each of the five generic PVA packages

|  | GAPPS ${ }^{9}$ | INMAT $^{10}$ | RAMAS <br> Metapop $^{11}$ | RAMAS <br> Stage $^{12}$ | VORTEX $^{13}$ |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Age structure | X | X | X | X | X |
| Stage structure |  |  | X | X |  |
| Survival and fecundity | X | X | X | X | X |
| Demographic stochasticity | X | X | X | X | X |
| Environmental variation | X | X | X | X | X |
| Inbreeding depression | X | X |  |  | X |
| Catastrophes | X |  | X | X | X |
| Breeding structure | X |  |  | X | X |
| Correlation in EV |  |  |  |  |  |
| No correlation in EV | X | X | X | X | X |
| Lognormal dist. of EV | X |  | X | X | X |
| Metapopulation structure |  |  | X | X |  |
| DD : Ceiling | X | X | X | X | X |
| DD: Logistic | X |  | X | X | X |
| DD: Beverton-Holt |  |  | X | X |  |
| DD: Ricker |  |  | X | X |  |
| DD: User defined function | X |  |  | X | X |

${ }^{\text {a }}$ Complete correlation in environmental stochasticity between survival and reproduction
${ }^{\mathrm{b}}$ Density dependence

## Table 2 Key population data for the 21 populations used in the retrospective PVA evaluations

| Class | Scientific name | Common name | \# Years ${ }^{\text {a }}$ | Est. set ${ }^{\text {b }}$ | Test set ${ }^{\text {c }}$ | Min $\mathrm{N}^{\text {d }}$ | Max N | $r^{e}$ | $V_{r}{ }^{\text {f }}$ | Ref ${ }^{9}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Aves | Copsychus sechellarum | Magpie robin | 17 | 1974-81 | 1981-90 | 20 | 46 | -0.030 | 0.024 | 14 |
|  | Grus americana | Whooping crane | 57 | 1939-68 | 1968-96 | 16 | 155 | 0.038 | 0.023 | 15 |
|  | Hirundo rustica | Danish swallow | 18 | 1971-79 | 1979-88 | 24 | 140 | -0.086 | 0.099 | 16 |
|  | Melospiza melodia | Song sparrow | 16 | 1975-83 | 1983-90 | 5 | 149 | -0.084 | 1.060 | 17 |
|  | Nannopterum harrisi | Galápagos cormorant | 21 | 1970-76 | 1976-91 | 84 | 150 | 0.022 | 0.040 | 18 |
|  | Parus atricapillus | Black-capped chickadee | 22 | 1960-71 | 1971-82 | 85 | 328 | 0.002 | 0.130 | 19 |
|  | Tricholimnas sylvestris | Lord Howe Is. woodhen | 18 | 1980-89 | 1989-97 | 26 | 223 | 0.057 | 0.016 | 20 |
|  | Zosterops lateralis | Heron Island silvereye | 27 | 1967-79 | 1979-93 | 225 | 445 | 0.019 | 0.076 | 21 |
| Mammalia | Canis lupus | Isle Royale gray wolf | 39 | 1959-78 | 1978-97 | 12 | 50 | 0.005 | 0.054 | 22 |
|  | Capra capra | Rhum goat | 35 | 1960-78 | 1978-95 | 66 | 175 | -0.130 | 0.103 | 23 |
|  | Cervas elaphus | Rhum deer (culled) ${ }^{\text {h }}$ | 14 | 1957-64 | 1964-71 | 149 | 348 | -0.070 | 0.042 | 24 |
|  | Cervas elaphus | Rhum deer (unculled) ${ }^{\text {h }}$ | 18 | 1972-81 | 1981-90 | 227 | 355 | 0.015 | 0.012 | 25 |
|  | Cynomys ludovicianus | Prairie dog | 13 | 1976-82 | 1982-89 | 92 | 143 | -0.007 | 0.023 | 26 |
|  | Lycaon pictus | Cape hunting dog | 22 | 1970-81 | 1981-92 | 0 | 77 | -0.021 | 0.431 | 27 |
|  | Marmota flaviensis | Yellow-bellied marmot | 31 | 1962-77 | 1977-93 | 18 | 116 | 0.047 | 0.068 | 28 |
|  | Odocoileus virginianus | White-tailed deer | 18 | 1952-61 | 1961-70 | 46 | 90 | 0.042 | 0.066 | 29 |
|  | Ovis aries | Soay sheep | 33 | 1960-67 | 1967-93 | 130 | 311 | 0.026 | 0.170 | 30 |
|  | Ovis aries | Boreray sheep ${ }^{\text {i }}$ | 40 | 1956-65 | 1965-96 | 172 | 699 | 0.107 | 0.095 | 30 |
|  | Ursus arctos horribilus | Grizzly bear | 38 | 1959-78 | 1978-97 | 120 | 296 | 0.017 | 0.014 | 31 |
| Pisces | Salvelinus fontinalis | Brook trout | 14 | 1949-55 | 1955-62 | 5258 | 8842 | 0.004 | 0.035 | 32 |
| Reptilia | Sceloporus graciosus | Sage-brush lizard | 11 | 1973-79 | 1969-79 | 70 | 201 | 0.097 | 0.131 | 33 |

${ }^{\text {a }}$ Total length of monitoring. ${ }^{\mathrm{b}}$ Time-period used to parameterise the PVA models. ${ }^{\mathrm{c}}$ Data used in the retrospective testing of the PVA predictions. ${ }^{d} N$ is population size. ${ }^{\ominus}$ Intrinsic rate of population increase estimated from the population time-series. ${ }^{\text {F }}$ Variance of $r$. ${ }^{9}$ Primary source of population data (most secondary sources are cited in the primary publication). ${ }^{\text {h }}$ Two time-periods from the same population were modelled separately, one involving a culled population and the other following the cessation of yearly culling. 'Soay sheep model used, as the Boreray is a different population of the same species facing similar ecological and environmental conditions on a nearby island.

Table 3 Key life-history data for the 21 populations used in the retrospective PVA evaluations

| Name | Breeding system ${ }^{\text {a }}$ | Age $1^{\text {st }}$ breeding ${ }^{\text {b }}$ | Fecundity $(C V)^{c}$ | Juvenile Survival (CV) ${ }^{\text {c }}$ | Adult <br> Survival (CV) ${ }^{\text {C }}$ | No. age classes | Gen <br> length ${ }^{\text {b }}$ | Max. age ${ }^{\text {b }}$ | Inbreeding modelled? | Density dependence |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Magpie robin | M | 1 | 0.334 (0.35) | 0.667 (0.09) | 0.667 (0.09) | 2 | 4.3 | 14 | Yes | Ceiling |
| Whooping crane | M | 4 | 0.268 (0.48) | 0.912 (0.12) | 0.912 (0.12)! | 5 | 10.0 | 50 | Yes | None |
| Danish swallow | M | 1 | 2.14 (0.13) | 0.299 (0.55) | 0.299 (0.55) | 2 | 2.4 | 10 | No | None |
| Song sparrow | M | 1 | 0.503 (0.43)* | 0.430 (0.74) | 0.301 (0.87)*! | 4 | 1.4 | 4 | Yes | Ceiling |
| Galápagos cormorant | M | 2 | 0.161 (0.66) | 0.567 (0.43) | 0.902 (0.06)* | 3 | 12.9 | 50 | Yes | Ceiling |
| Black-capped chickadee | M | 1 | 0.933 (0.71) | 0.433 (0.19) | 0.652 (0.17) | 7 | 3.3 | 7 | No | None |
| Lord Howe I. Woodhen | M | 1 | 0.938 (0.14)* | 0.348 (0.21) | 0.760 (0.17)! | 2 | 2.7 | 7 | Yes | Ceiling |
| Heron Island silvereye | M | 1 | 0.449 (0.65)* | 0.577 (0.15) | 0.624 (0.24)*! | 5 | 2.6 | 11 | Yes | Ceiling |
| Isle Royale gray wolf | A | 3 | 0.475 (0.32) | 0.819 (0.15) | 0.819 (0.15) | 4 | 8.1 | 14 | Yes | Ceiling |
| Rhum goat | P | 1 | 0.198 (0.45)* | 0.888 (0.37) | 0.792 (0.42)* | 10 | 4.6 | 10 | No | Ceiling |
| Rhum deer (culled) | P | 3 | 0.216 (0.48)* | 0.706 (0.45) | 0.668 (0.23)* | 17 | 6.4 | 17 | No | None |
| Rhum deer (unculled) | P | 3 | 0.193 (0.36)*\# | 0.645 (0.16) | 0.918 (0.13)* | 21 | 9.5 | 21 | No | Contest |
| Prairie dog | P | 1 | 0.381 (0.00)*\# | 0.519 (0.00) | 0.687 (0.00)* | 2 | 2.7 | 8 | No | Logistic |
| Cape hunting dog | M | 2 | 0.277 (0.87) | 0.200 (0.70) | 0.862 (0.07) | 2 | 6.0 | 10 | Yes | Ceiling |
| Yellow-bellied marmot | P | 2 | 0.624 (0.51)\# | 0.472 (0.38)\# | 0.786 (0.14)* | 2 | 5.2 | 15 | No | Contest |
| White-tailed deer | P | 1 | 0.497 (0.16) | 0.433 (0.00) | 0.607 (0.17)* | 3 | 2.2 | 12 | No | Ceiling |
| Soay sheep | P | 1 | 0.265 (0.35)* | 0.821 (0.05)\# | 0.821 (0.05)*\# | 2 | 4.2 | 12 | No | Scramble |
| Boreray sheep | P | 1 | 0.265 (0.35)* | 0.821 (0.05)\# | 0.821 (0.05)*\# | 2 | 4.2 | 12 | No | Scramble |
| Grizzly bear | P | 5 | 0.328 (0.25)* | 0.890 (0.11) | 0.920 (0.11)* | 5 | 15.8 | 25 | Yes | Ceiling |
| Brook trout | P | 1 | 2.116 (0.29)* | 0.397 (0.14) | 0.136 (0.21)* | 5 | 2.4 | 5 | No | None |
| Sage-brush lizard | P | 2 | 1.083 (0.52) | 0.439 (0.41) | 0.600 (0.29)* | 4 | 3.1 | 7 | No | None |

${ }^{\mathrm{a}} M=$ monogamous, $P=$ polygamous, $A=$ alpha pair. ${ }^{\mathrm{b}}$ Age of first breeding, generation length and maximum age are given in years. ${ }^{\mathrm{c}}$ Average fecundity, juvenile and adult survival rates (and their coefficient of variation, representing environmental stochasticity) are given. Detailed age-structure (indicated by *) or density dependent rates (\#) were used in the PVA models when sufficient data existed. Catastrophes were modelled in cases denoted by a "!" in the survival column.

Table 4 The relative rank and overall bias of each PVA package for each of the 21 populations

| Name | GAPPS | INMAT | R META R STAGE VORTEX |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| Magpie robin | $1-$ | $4-$ | $2-$ | $3-$ | $5-$ |
| Whooping crane | $4-$ | $5-$ | $2+$ | $3-$ | $1+$ |
| Danish swallow | $1+$ | $3-$ | $2+$ | $4-$ | $5-$ |
| Song sparrow | $3-$ | $5-$ | $4-$ | $1-$ | $2-$ |
| Galápagos cormorant | $1-$ | $4-$ | $2-$ | $3-$ | $5-$ |
| Black-capped chickadee | $5+$ | $2+$ | $3-$ | $4-$ | $1+$ |
| Lord Howe Is. woodhen | $2+$ | $4+$ | $3+$ | $5+$ | $1-$ |
| Heron Island silvereye | $2+$ | $1+$ | $4-$ | $5-$ | $3-$ |
| Isle Royale gray wolf | $1+$ | $4+$ | $5+$ | $3+$ | $2+$ |
| Rhum goat | $4-$ | $1-$ | $2-$ | $5-$ | $3-$ |
| Rhum deer (culled) | $5-$ | $1-$ | $3-$ | $2-$ | $4-$ |
| Rhum deer (unculled) | $3+$ | $5+$ | $1-$ | $4+$ | $2+$ |
| Prairie dog | $2+$ | $5+$ | $3+$ | $1+$ | $4+$ |
| Cape hunting dog | $2-$ | $3-$ | $1-$ | $5-$ | $4-$ |
| Yellow-bellied marmot | $4-$ | $5-$ | $1-$ | $2-$ | $3-$ |
| White-tailed deer | $5+$ | $3+$ | $1+$ | $4+$ | $2+$ |
| Soay sheep | $2+$ | $5-$ | $4+$ | $1+$ | $3+$ |
| Boreray sheep | $2+$ | $5-$ | $4+$ | $1+$ | $3+$ |
| Grizzly bear | $2-$ | $5-$ | $3-$ | $4-$ | $1-$ |
| Brook trout | N/A | $3+$ | $2+$ | $4+$ | $1-$ |
| Sage-brush lizard | $5+$ | $3+$ | $1+$ | $4+$ | $2+$ |
| Average rank | 2.8 | 3.6 | 2.5 | 3.2 | 2.7 |

Rank was based on the goodness of fit to actual population size - a least squares $\left(\chi^{2}=\Sigma(P-A)^{2} / \mathrm{A}\right)$ estimate, where $P$ is the predicted median size at each time-step and $A$ is the actual population size. Ranks do not differ across packages ( $\mathrm{P}=0.097$ ). To be identical, all packages would have a rank of 3 $([1+2+3+4+5] / 5)$. The bias represents the sum of the difference between predicted and observed size at each time point, averaged over 500 simulation replicates. A negative bias ( - ) indicates an underestimation of population size, a positive bias (+) the converse.

