Maximum Likelihood in Phylogenetics

26 January 2011

Workshop on Molecular Evolution
Český Krumlov, Česká republika

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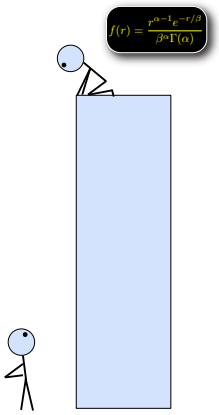
Goals

Explain jargon
Increase comfort level
Provide background
In other words...give a hand up



The Plan

- Probability review
  - The AND and OR rules
  - Independence of events
- Likelihood
  - What does it mean?
  - Likelihood of a single sequence
  - Maximum likelihood distances
  - Likelihoods of trees
- Substitution models
  - Markov model basics
  - Transition probabilities
  - Survey of models
  - Rate heterogeneity
  - Codon models

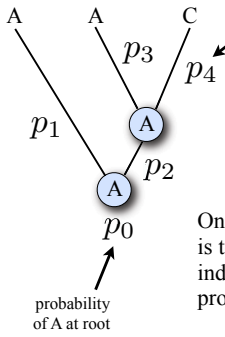
Combining probabilities

- *Multiply* probabilities if the component events must happen **simultaneously** (i.e. where you would naturally use the word AND when describing the problem)

Using 2 dice, what is the probability of

$(1/6) \times (1/6) = 1/36$

AND rule in phylogenetics



probability of C at end of branch given A at beginning

we have observed these states

pretend (for now) we also know these ancestral states

$$\Pr(AAC, AA) = p_0\ p_1\ p_2\ p_3\ p_4$$

One use of the AND rule in phylogenetics is to combine probabilities associated with individual branches to produce the overall probability of the data for one site.

Combining probabilities

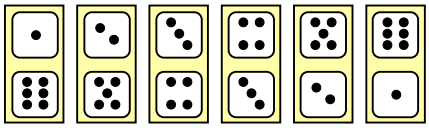
- *Add* probabilities if the component events are **mutually exclusive** (i.e. where you would naturally use the word OR in describing the problem)

Using one die, what is the probability of

$(1/6) + (1/6) = 1/3$

Combining AND and OR

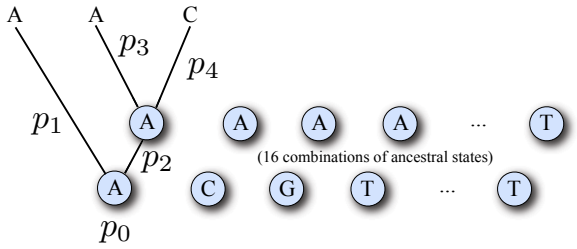
What is the probability that the sum of two dice is 7?



(1/36) + (1/36) + (1/36) + (1/36) + (1/36) + (1/36) = 1/6

1 and 6 | 2 and 5 | 3 and 4 | 4 and 3 | 5 and 2 | 6 and 1

Using both AND and OR in phylogenetics



AND rule used to compute probability of the observed data for *each combination* of ancestral states.

OR rule used to combine different combinations of ancestral states.

Independence

This is always true...

$$\Pr(\text{A and B}) = \Pr(\text{A})\,\Pr(\text{B}|\text{A})$$

joint probability | conditional probability

If we can say this...

$$\Pr(\text{B}|\text{A}) = \Pr(\text{B})$$

...then events A and B are **independent** and we can express the joint probability as the product of Pr(A) and Pr(B)

$$\Pr(\text{A and B}) = \Pr(\text{A})\,\Pr(\text{B})$$

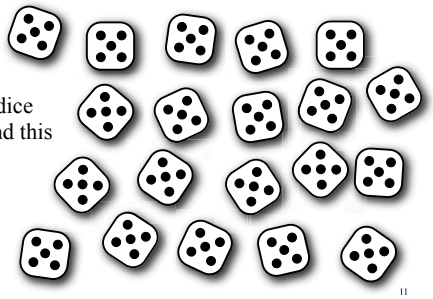
Likelihood

The Likelihood Criterion

The probability of the observations computed using a model tells us how surprised we should be.

The preferred model is the one that surprises us least.

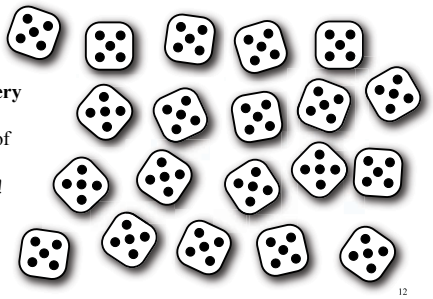
Suppose I threw 20 dice down on the table and this was the result...



The Fair Dice model

$$\Pr(\text{obs.}|\text{fair dice model}) = \left(\frac{1}{6}\right)^{20} = \frac{1}{3,656,158,440,062,976}$$

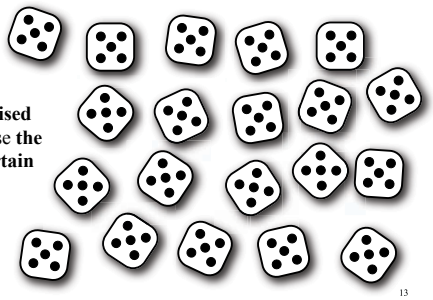
You should have been **very surprised** at this result because the probability of this event is **very small**: only 1 in 3.6 quadrillion!



The Trick Dice model

(assumes dice each have 5 on every side)

$$\Pr(\text{obs.}|\text{trick dice model}) = 1^{20} = 1$$



You should **not be surprised at all** at this result because **the observed outcome is certain** under this model

Results

Model	Likelihood	Surprise level
Fair Dice	$\frac{1}{3{,}656{,}158{,}440{,}062{,}976}$	Very, *very*, ***very*** surprised
Trick Dice	1.0	Not surprised at all

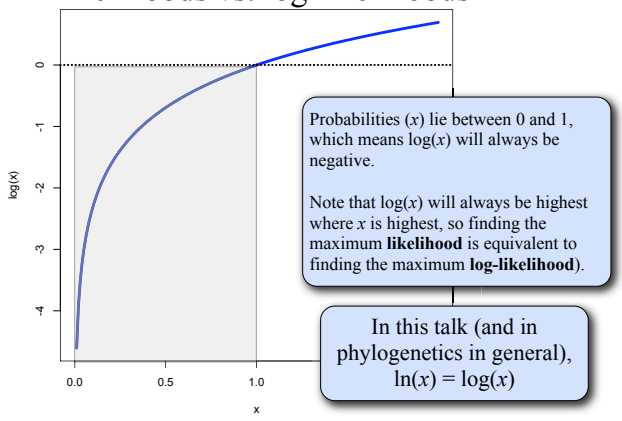
winning model maximizes likelihood (and thus minimizes surprise)

Likelihood and model comparison

- Analyses using likelihoods ultimately involve **model comparison**
- The models compared can be **discrete** (as in the fair vs. trick dice example)
- More often the models compared differ **continuously**:
  - Model 1: branch length is 0.05
  - Model 2: branch length is 0.06

Rather than having an infinity of models, we instead think of the branch length as a **parameter** within one model

Likelihoods vs. log-likelihoods



Probabilities (x) lie between 0 and 1, which means $\log(x)$ will always be negative.

Note that $\log(x)$ will always be highest where x is highest, so finding the maximum **likelihood** is equivalent to finding the maximum **log-likelihood**).

In this talk (and in phylogenetics in general), $\ln(x) = \log(x)$

Likelihood of a single sequence

First 32 nucleotides of the ψη-globin gene of gorilla:

GAAGTCCTTGAGAAATAAACTGCACACACTGG

$$L = \pi_G\pi_A\pi_A\pi_G\pi_T\pi_C\pi_C\pi_T\pi_T\pi_G\pi_A\pi_G\pi_A\pi_A\pi_A\pi_T\pi_A\pi_A\pi_A\pi_C\pi_T\pi_G\pi_C\pi_A\pi_C\pi_A\pi_C\pi_A\pi_C\pi_T\pi_G\pi_G$$
$$= \pi_A^{12}\pi_C^{7}\pi_G^{7}\pi_T^{6}$$

Note that we are assuming independence between sites here

$$\log L = 12\log(\pi_A) + 7\log(\pi_C) + 7\log(\pi_G) + 6\log(\pi_T)$$

We can already see by eye-balling this that a model (e.g. F81) allowing **unequal** base frequencies will **fit better** than a model that assumes **equal** base frequencies (e.g. JC69) because there are about twice as many As as there are Cs, Gs and Ts.

Likelihood ratio test

Likelihood ratio tests can be used to evaluate whether an **unconstrained** model fits the data significantly better than a **constrained** version of the same model.

Find *maximum* logL under F81 (unconstrained) model:

$$\begin{aligned}\log L &= 12\log(\pi_A) + 7\log(\pi_C) + 7\log(\pi_G) + 6\log(\pi_T)\\ &= 12\log(0.375) + 7\log(0.219) + 7\log(0.219) + 6\log(0.187)\\ &= -43.1\end{aligned}$$

Find *maximum* logL under JC69 (constrained) model:

$$\begin{aligned}\log L &= 12\log(\pi_A) + 7\log(\pi_C) + 7\log(\pi_G) + 6\log(\pi_T)\\ &= 12\log(0.25) + 7\log(0.25) + 7\log(0.25) + 6\log(0.25)\\ &= -44.4\end{aligned}$$

Likelihood ratio test

Calculate the likelihood ratio test statistic:

$$R = -2\Big[\log(L_{\text{JC69}}) - \log(L_{\text{F81}})\Big]$$
$$= -2\Big[-44.4 - (-43.1)\Big] = 2.6$$

(Note that the log-likelihoods used in the test statistic have been *maximized* under each model separately)

F81 does fit better (-43.1 > -44.4), but not significantly better (P = 0.457, chi-squared with 3 d.f.*)

*The number of degrees of freedom equals the difference between the two models in the number of free parameters. In this case, F81 has 3 parameters and JC69 has 0, so d.f. = 3 – 0 = 3

Likelihood of the simplest tree

sequence 1 ——— sequence 2

To keep things simple, assume that the sequences are only 2 nucleotides long:

$$L = L_1\, L_2$$
$$= \left[\left(\frac{1}{4}\right)\left(\frac{1}{4}+\frac{3}{4}e^{-4\alpha t}\right)\right]\left[\left(\frac{1}{4}\right)\left(\frac{1}{4}-\frac{1}{4}e^{-4\alpha t}\right)\right]$$

Pr(G) | Pr(G|G, α*t*) | Pr(A) | Pr(G|A, α*t*)

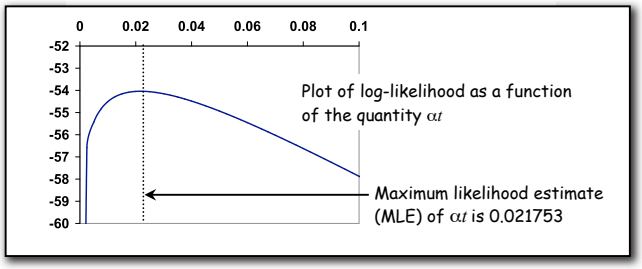
Note that we are NOT assuming independence here

Maximum likelihood estimation

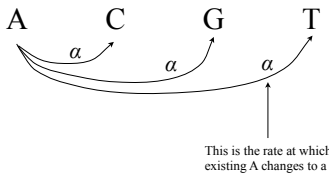
First 32 nucleotides of the ψη-globin gene of gorilla and orangutan:

```
gorilla   GAAGTCCTTGAGAAATAAACTGCACACACTGG
orangutan GGACTCCTTGAGAAATAAACTGCACACACTGG
```

$$L = \left[\left(\tfrac{1}{4}\right)\left(\tfrac{1}{4}+\tfrac{3}{4}e^{-4\alpha t}\right)\right]^{30}\left[\left(\tfrac{1}{4}\right)\left(\tfrac{1}{4}-\tfrac{1}{4}e^{-4\alpha t}\right)\right]^{2}$$



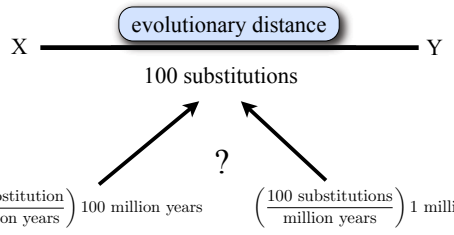
number of substitutions = rate × time



Overall substitution rate is 3α, so the expected number of substitutions (v) is

$$v = 3\alpha t$$

Rate and time are confounded



Andrew will discuss (next week) models in which constraints on times can be used to infer rates (and vice versa), but without some extra information or constraints, sequence data allow only estimation of the **number** of substitutions.

A convenient convention

Because rate and time are confounded, it is convenient to arbitrarily standardize things by setting the rate to a value such that **one substitution** is expected to occur in **one unit of time** for each site.

This results in "time" (the length of a branch) being measured in units of **evolutionary distance (expected number of substitutions per site)** rather than years (or some other calendar unit).

evolutionary distance $v = 3\alpha t$

$$v = 3\left(\frac{1}{3}\right)t$$

Setting α=1/3 results in v equalling t

Evolutionary distances for several common models

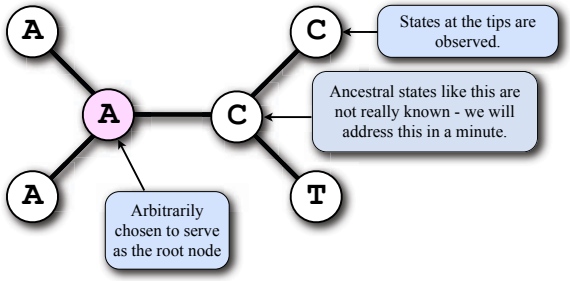
Model	Expected no. substitutions: $v = \{r\}t$
JC69	$v = \{3\alpha\}\, t$
F81	$v = \{2\mu(\pi_R\pi_Y + \pi_A\pi_G + \pi_C\pi_T)\}\, t$
K80	$v = \{\beta(\kappa + 2)\}\, t$
HKY85	$v = \{2\mu\,[\pi_R\pi_Y + \kappa(\pi_A\pi_G + \pi_C\pi_T)]\}\, t$

In the formulas above, the overall rate r (in curly brackets) is a function of all parameters in the substitution model.

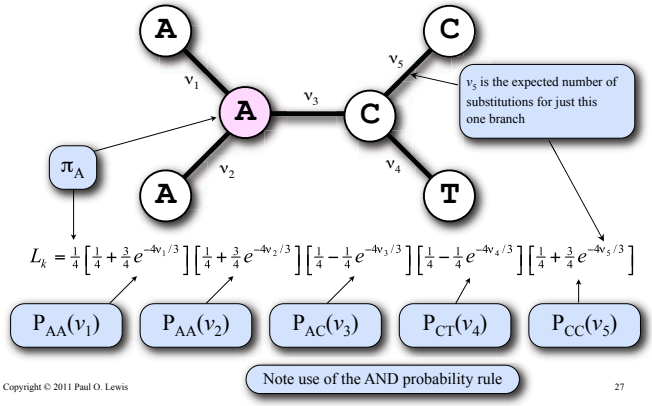
One of the parameters of the substitution model can always be determined from the branch length (using our convention that $v = t$).

Likelihood of an unrooted tree

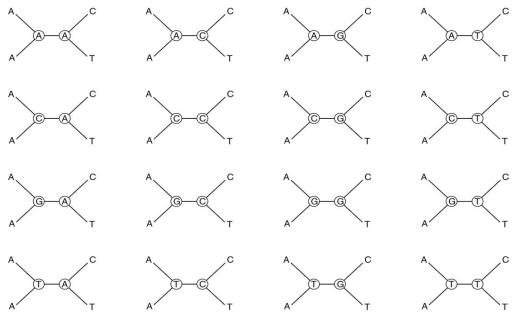
(data shown for only one site)



Likelihood for site *k*



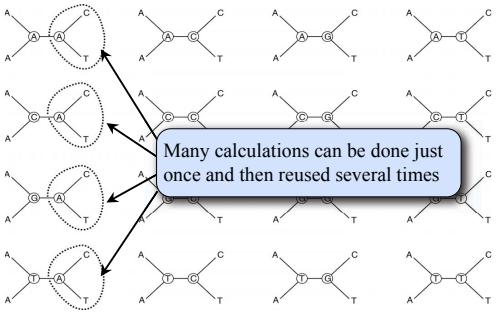
Brute force approach would be to calculate L_k for all 16 combinations of ancestral states and sum them



Note use of the OR probability rule

Pruning algorithm

(same result, less time)



Felsenstein, J. 1981. Evolutionary trees from DNA sequences: a maximum likelihood approach. *Journal of Molecular Evolution* **17**:368-376

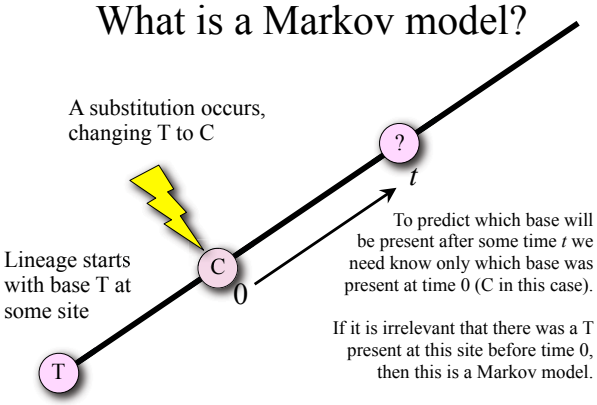
Substitution Models

Jukes-Cantor (JC69) model

- The four bases (A, C, G, T) are expected to be **equally frequent** in sequences ($\pi_A = \pi_C = \pi_G = \pi_T = 0.25$)
- Assumes **same rate** for all types of substitution ($r_{A\to C} = r_{A\to G} = r_{A\to T} = r_{C\to G} = r_{C\to T} = r_{G\to T} = \alpha$)
- Usually described as a **1-parameter** model (the parameter being the branch length)
  - Remember, however, that each branch in a tree can have its own length, so there are really as many parameters in the model as there are edges in the tree!
- Assumes substitution is a **Markov** process...

Jukes, T. H., and C. R. Cantor. 1969. Evolution of protein molecules. Pages 21-132 *in* H. N. Munro (ed.), *Mammalian Protein Metabolism*. Academic Press, New York.

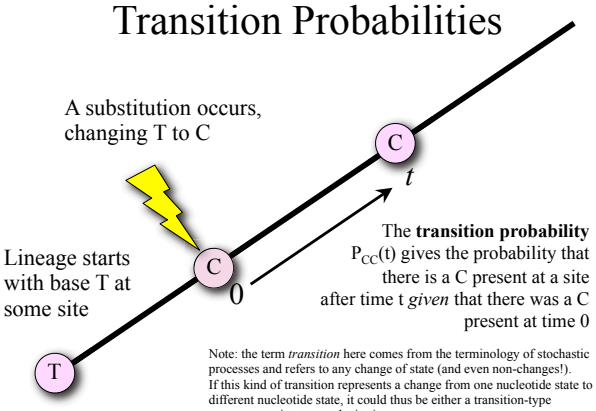
What is a Markov model?



To predict which base will be present after some time *t* we need know only which base was present at time 0 (C in this case).

If it is irrelevant that there was a T present at this site before time 0, then this is a Markov model.

Transition Probabilities



The **transition probability** $P_{CC}(t)$ gives the probability that there is a C present at a site after time t *given* that there was a C present at time 0

Note: the term *transition* here comes from the terminology of stochastic processes and refers to any change of state (and even non-changes!). If this kind of transition represents a change from one nucleotide state to a different nucleotide state, it could thus be either a transition-type or a transversion-type substitution.

Jukes-Cantor transition probabilities

Here is the probability that a site starting in state T will end up in state G after time *t* when the individual substitution rates are all α:

$$P_{TG}(t) = \frac{1}{4}\left(1 - e^{-4\alpha t}\right)$$

The JC69 model has only one unknown quantity: αt

(The symbol *e* represents the base of the natural logarithms: its value is 2.718281828459045...)

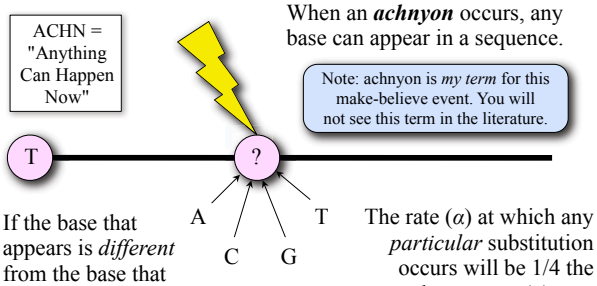
Where does a transition probability formula such as this come from?

"ACHNyons" vs. substitutions

ACHN = "Anything Can Happen Now"

When an ***achnyon*** occurs, any base can appear in a sequence.

Note: achnyon is *my term* for this make-believe event. You will not see this term in the literature.



If the base that appears is *different* from the base that was already there, then a **substitution** event has occurred.

The rate (α) at which any *particular* substitution occurs will be 1/4 the achnyon rate (μ). That is, $\alpha = \mu/4$ (or $\mu = 4\alpha$)

Deriving a transition probability

Calculate the probability that a site currently T will change to G over time *t* when the rate of this particular substitution is α:

Pr(zero achnyons) = $e^{-\mu t}$ (Poisson probability of zero events)

Pr(at least 1 achnyon) = $1 - e^{-\mu t}$

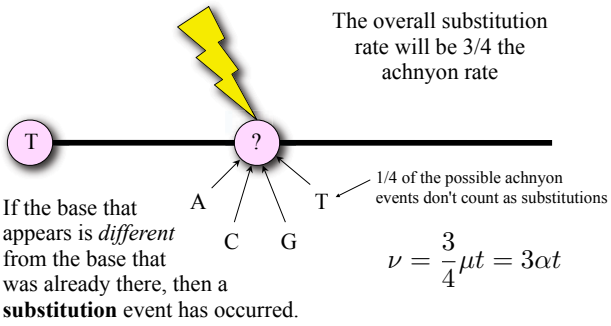
Pr(last achnyon results in base G) = $\frac{1}{4}$

Pr(end in G | start in T) = $\frac{1}{4}\left(1 - e^{-\mu t}\right)$

Remember that the rate (α) of any particular substitution is one fourth the achnyon rate (μ):

$$P_{GT}(t) = \frac{1}{4}\left(1 - e^{-4\alpha t}\right)$$

Expected number of substitutions



The overall substitution rate will be 3/4 the achnyon rate

1/4 of the possible achnyon events don't count as substitutions

If the base that appears is *different* from the base that was already there, then a **substitution** event has occurred.

$$\nu = \frac{3}{4}\mu t = 3\alpha t$$

Transition Probabilities: Remarks

$$\begin{aligned} P_{TA}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TC}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TG}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TT}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ \hline & \quad 1 - e^{-4\alpha t} \end{aligned}$$

These should add to 1.0 because T *must* change to something!

Doh! Something must be wrong here...

Transition Probabilities: Remarks

$$\begin{aligned} P_{TA}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TC}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TG}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) \\ P_{TT}(t) &= \frac{1}{4}(1 - e^{-4\alpha t}) + e^{-4\alpha t} \end{aligned}$$

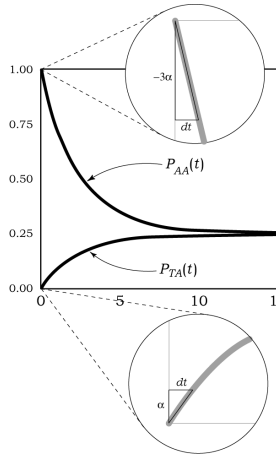
Forgot to account for the possibility of *no* achnyons over time *t*

Equilibrium frequencies

- The JC69 model assumes that the frequencies of the four bases (A, C, G, T) are equal
- The equilibrium relative frequency of each base is thus 0.25
- Why are they called *equilibrium* frequencies?

Equilibrium Frequency Simulation

Pr(A|A) and Pr(A|T) as a function of time



Upper curve assumes we started with A at time 0. Over time, the probability of still seeing an A at this site **drops** because rate of changing to one of the other three bases is 3α (so rate of staying the same is -3α).

The equilibrium relative frequency of A is 0.25

Lower curve assumes we started with some state *other* than A (T is used here). Over time, the probability of seeing an A at this site **grows** because the rate at which the current base will change into an A is α.

JC69 rate matrix

1 parameter:
α

From \ To	A	C	G	T
A	-3α	α	α	α
C	α	-3α	α	α
G	α	α	-3α	α
T	α	α	α	-3α

Jukes, T. H., and C. R. Cantor. 1969. Evolution of protein molecules. Pages 21-132 *in* H. N. Munro (ed.), *Mammalian Protein Metabolism*. Academic Press, New York.

K80 (or K2P) rate matrix

2 parameters:
α
β

From \ To	A	C	G	T
A	$-\alpha-2\beta$	β	α	β
C	β	$-\alpha-2\beta$	β	α
G	α	β	$-\alpha-2\beta$	β
T	β	α	β	$-\alpha-2\beta$

transition rate

transversion rate

Kimura, M. 1980. A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. Journal of Molecular Evolution 16:111-120.

K80 rate matrix
(looks different, but actually the same)

2 parameters:
κ
β

	A	C	G	T
A	$-\beta(\kappa+2)$	β	$\kappa\beta$	β
C	β	$-\beta(\kappa+2)$	β	$\kappa\beta$
G	$\kappa\beta$	β	$-\beta(\kappa+2)$	β
T	β	$\kappa\beta$	β	$-\beta(\kappa+2)$

All I've done is re-parameterize the rate matrix, letting κ equal the *transition/transversion rate ratio* $\longrightarrow$ $\kappa = \frac{\alpha}{\beta}$

Note: the K80 model is identical to the JC69 model if $\kappa = 1$ ($\alpha = \beta$)

F81 rate matrix

4 parameters:
μ
π_A
π_C
π_G

	A	C	G	T
A	$-\mu(1-\pi_A)$	$\pi_C\mu$	$\pi_G\mu$	$\pi_T\mu$
C	$\pi_A\mu$	$-\mu(1-\pi_C)$	$\pi_G\mu$	$\pi_T\mu$
G	$\pi_A\mu$	$\pi_C\mu$	$-\mu(1-\pi_G)$	$\pi_T\mu$
T	$\pi_A\mu$	$\pi_C\mu$	$\pi_G\mu$	$-\mu(1-\pi_T)$

Note: the F81 model is identical to the JC69 model if all base frequencies are equal

Felsenstein, J. 1981. Evolutionary trees from DNA sequences: a maximum likelihood approach. Journal of Molecular Evolution 17:368-376.

HKY85 rate matrix

5 parameters:
κ
β
π_A
π_C
π_G

	A	C	G	T
A	—	$\pi_C\beta$	$\pi_G\beta\kappa$	$\pi_T\beta$
C	$\pi_A\beta$	—	$\pi_G\beta$	$\pi_T\beta\kappa$
G	$\pi_A\beta\kappa$	$\pi_C\beta$	—	$\pi_T\beta$
T	$\pi_A\beta$	$\pi_C\beta\kappa$	$\pi_G\beta$	—

A dash means equal to negative sum of other elements on the same row

Note: the HKY85 model is identical to the F81 model if $\kappa = 1$. If, in addition, all base frequencies are equal, it is identical to JC69.

Hasegawa, M., H. Kishino, and T. Yano. 1985. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. Journal of Molecular Evolution 21:160-174.

F84 vs. HKY85

F84 model:

- μ rate of process generating *all types of substitutions*
- $k\mu$ rate of process generating *only transitions*

Becomes F81 model if $k = 0$

HKY85 model:

- β rate of process generating *only transversions*
- $\kappa\beta$ rate of process generating *only transitions*

Becomes F81 model if $\kappa = 1$

F84 first used in Felsenstein's PHYLIP package in 1984, first published by: Kishino, H., and M. Hasegawa. 1989. Evaluation of the maximum likelihood estimate of the evolutionary tree topologies from DNA sequence data, and the branching order in hominoidea. Journal of Molecular Evolution 29: 170-179.

GTR rate matrix

	A	C	G	T
A	$-$	$\pi_C a\mu$	$\pi_G b\mu$	$\pi_T c\mu$
C	$\pi_A a\mu$	$-$	$\pi_G d\mu$	$\pi_T e\mu$
G	$\pi_A b\mu$	$\pi_C d\mu$	$-$	$\pi_T f\mu$
T	$\pi_A c\mu$	$\pi_C e\mu$	$\pi_G f\mu$	$-$

9 parameters:
π_A
π_C
π_G
a
b
c
d
e
μ

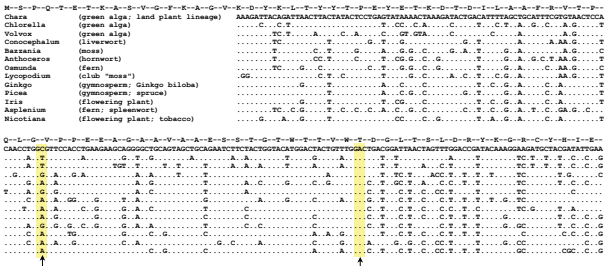
Identical to the F81 model if $a = b = c = d = e = f = 1$. If, in addition, all the base frequencies are equal, GTR is identical to JC69. If $a = c = d = f = \beta$ and $b = e = \kappa\beta$, GTR becomes the HKY85 model.

Lanave, C., G. Preparata, C. Saccone, and G. Serio. 1984. A new method for calculating evolutionary substitution rates. Journal of Molecular Evolution 20:86-93.

Rate Heterogeneity

Green Plant rbc*L*

First 88 amino acids (translation is for *Zea mays*)



All four bases are observed at some sites...

...while at other sites, only one base is observed

Site-specific rates

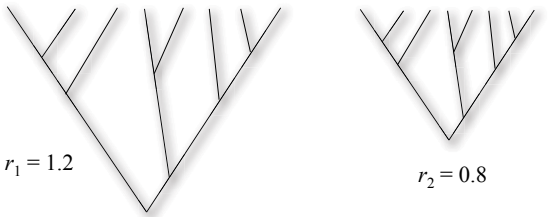
Each defined subset (e.g. gene, codon position) has its own relative rate

r_1 applies to subset 1
(e.g. sites 1 - 1000)

r_2 applies to subset 2
(e.g. sites 1001-2000)

Site-specific rates

$$L = \underbrace{\Pr(D_1|r_1)\cdots\Pr(D_{1000}|r_1)}_{\text{Gene 1}}\ \underbrace{\Pr(D_{1001}|r_2)\cdots\Pr(D_{2000}|r_2)}_{\text{Gene 2}}$$



Site-specific rates

JC69 transition probabilities that would be used for every site if rate *homo*geneity were assumed:

$$P_{ii}(t) = \frac{1}{4} + \frac{3}{4}e^{-4\alpha t}$$

$$P_{ij}(t) = \frac{1}{4} - \frac{1}{4}e^{-4\alpha t}$$

Site specific rates

JC69 transition probabilities that would be used for sites in **gene 1**:

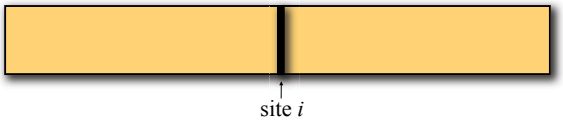
$$P_{ii}(t) = \frac{1}{4} + \frac{3}{4}e^{-4r_1\alpha t}$$
$$P_{ij}(t) = \frac{1}{4} - \frac{1}{4}e^{-4r_1\alpha t}$$

JC69 transition probabilities that would be used for sites in **gene 2**:

$$P_{ii}(t) = \frac{1}{4} + \frac{3}{4}e^{-4r_2\alpha t}$$
$$P_{ij}(t) = \frac{1}{4} - \frac{1}{4}e^{-4r_2\alpha t}$$

Mixture Models

All relative rates applied to every site

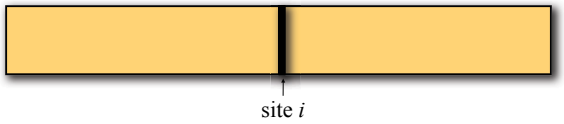


$$L_i = \Pr(D_i|r_1)\Pr(r_1) + \Pr(D_i|r_2)\Pr(r_2)$$

Common examples { Invariable sites (I) model; Discrete Gamma (G) model

Invariable Sites Model

A fraction p_{invar} of sites are assumed to be invariable (i.e. rate = 0.0)



$$L_i = \Pr(D_i|r_1)p_{\text{invar}} + \Pr(D_i|r_2)(1 - p_{\text{invar}})$$

$r_1 = 0.0$

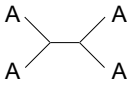
$r_2 = \dfrac{1}{1 - p_{\text{invar}}}$

Allows for the possibility that any given site could be variable or invariable

Reeves, J. H. 1992. Heterogeneity in the substitution process of amino acid sites of proteins coded for by mitochondrial DNA. Journal of Molecular Evolution 35:17-31.

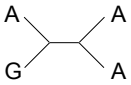
Invariable sites model

If site *i* is a *constant* site, both terms will contribute to the site likelihood:



$$L_i = \Pr(D_i|0.0)p_{\text{invar}} + \Pr(D_i|r_2)(1 - p_{\text{invar}})$$

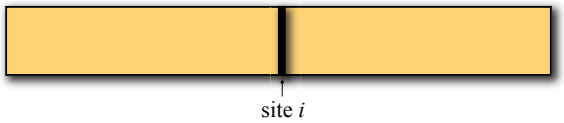
If site *i* is a *variable* site, there is no way to explain the data with a zero rate, so the first term is zero:



$$L_i = \cancel{\Pr(D_i|0.0)p_{\text{invar}}} + \Pr(D_i|r_2)(1 - p_{\text{invar}})$$

Discrete Gamma Model

No relative rates are exactly 0.0, and all are equally probable



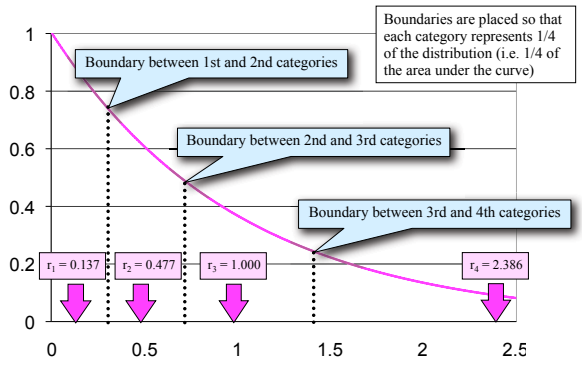
$$L = (\tfrac{1}{4})\Pr(D_i|r_1) + (\tfrac{1}{4})\Pr(D_i|r_2) + (\tfrac{1}{4})\Pr(D_i|r_3) + (\tfrac{1}{4})\Pr(D_i|r_4)$$

Relative rates are constrained to a discrete gamma distribution
Number of rate categories can vary (4 used here)

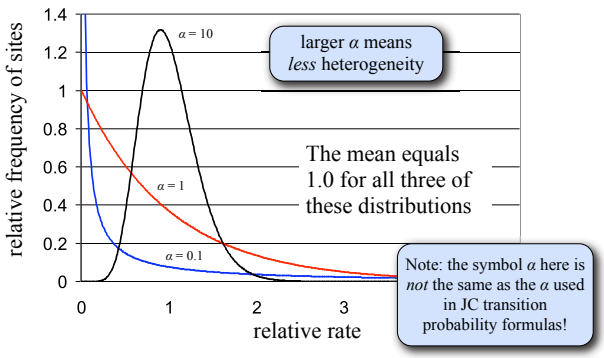
Yang, Z. 1993. Maximum-likelihood estimation of phylogeny from DNA sequences when substitution rates differ over sites. Molecular Biology and Evolution 10:1396-1401.

Yang, Z. 1994. Maximum likelihood phylogenetic estimation from DNA sequences with variable rates over sites: approximate methods. Journal of Molecular Evolution 39:306-314.

Relative rates in 4-category case



Gamma distributions



Codon models

The Genetic Code

First 12 nucleotides at the 5' end of the *rbc*L gene in corn:

```
5'-ATG|TCA|CCA|CAA-3'   coding strand
3'-TAC|AGT|GGT|GTT-5'   template strand
```

} DNA double helix

transcription

```
5'-AUG|UCA|CCA|CAA-3'   mRNA
```

translation

```
N-Met|Ser|Pro|Gln-C     polypeptide
```

Codon models treat codons as the independent units, not individual nucleotide sites.

Genetic Code

	U	C	A	G	
U	UUU Phe	UCU Ser	UAU Tyr	UGU Cys	U
	UUC Phe	UCC Ser	UAC Tyr	UGC Cys	C
	UUA Leu	UCA Ser	UAA Stop	UGA Stop	A
	UUG Leu	UCG Ser	UAG Stop	UGG Trp	G
C	CUU Leu	CCU Pro	CAU His	CGU Arg	U
	CUC Leu	CCC Pro	CAC His	CGC Arg	C
	CUA Leu	CCA Pro	CAA Gln	CGA Arg	A
	CUG Leu	CCG Pro	CAG Gln	CGG Arg	G
A	AUU Ile	ACU Thr	AAU Asn	AGU Ser	U
	AUC Ile	ACC Thr	AAC Asn	AGC Ser	C
	AUA Ile	ACA Thr	AAA Lys	AGA Arg	A
	AUG Met	ACG Thr	AAG Lys	AGG Arg	G
G	GUU Val	GCU Ala	GAU Asp	GGU Gly	U
	GUC Val	GCC Ala	GAC Asp	GGC Gly	C
	GUA Val	GCA Ala	GAA Glu	GGA Gly	A
	GUG Val	GCG Ala	GAG Glu	GGG Gly	G

http://www.langara.bc.ca/biology/mario/Assets/Geneticode.jpg

First codon models

- Muse and Gaut model (MG94) is simplest
  - α = synonymous substitution rate
  - β = nonsynonymous substitution rate
  - π_A, π_C, π_G, π_T = base frequencies
- Goldman and Yang model (GY94) similar
  - accounts for synon./nonsynon. *and* trs/trv bias *and* amino acid properties (later simplified, see Yang et al. 1998)

Muse, S. V., and B. S. Gaut. 1994. A likelihood approach for comparing synonymous and nonsynonymous substitution rates, with application to the chloroplast genome. Molecular Biology and Evolution 11:715-724.

Goldman, N., and Z. Yang. 1994. A codon-based model of nucleotide substitution for protein-coding DNA sequences. Molecular Biology and Evolution 11:725-736.

Yang, Z., Nielsen, R., and Hasegawa, M. 1998. Models of amino acid substitution and applications to mitochondrial protein evolution. Molecular Biology and Evolution 15:1600-1611.

Table I. Part of Muse and Gaut's 61 × 61 instantaneous rate matrix[a]

Codon before substitution (the 'from' state)	Codon after substitution (the 'to' state)							
	TTT (Phe)	TTC (Phe)	TTA (Leu)	TTG (Leu)	CTT (Leu)	CTC (Leu)	...	GGG (Gly)
TTT (Phe)	---	$\alpha\pi_C$	$\beta\pi_A$	$\beta\pi_G$	$\beta\pi_C$	0	...	0
TTC (Phe)	$\alpha\pi_T$	---	$\beta\pi_A$	$\beta\pi_G$	0	$\beta\pi_C$	...	0
TTA (Leu)	$\beta\pi_T$	$\beta\pi_C$	---	$\alpha\pi_G$	0	0	...	0
TTG (Leu)	$\beta\pi_T$	$\beta\pi_C$	$\alpha\pi_A$	---	0	0	...	0
CTT (Leu)	$\beta\pi_T$	0	0	0	---	$\alpha\pi_C$	...	0
CTC (Leu)	0	$\beta\pi_T$	0	0	$\alpha\pi_T$	---	...	0
⋮	⋮	⋮	⋮	⋮	⋮	⋮	⋱	⋮
GGG (Gly)	0	0	0	0	0	0	...	---

Note that it is still easy for the change CTT → TTA to occur, it just requires more than one instant of time

Instantaneous rate is 0.0 if two or more nucleotides must change during the codon transition

Table 1 from: Lewis, P. O. 2001. Phylogenetic systematics turns over a new leaf. Trends in Ecology and Evolution 16:30-37.

Interpreting codon model results

$\omega = \beta/\alpha$ is the nonsynonymous/synonymous rate ratio

omega	mode of selection	example(s)
$\omega < 1$	stabilizing selection (nucleotide substitutions rarely change the amino acid)	functional protein coding genes
$\omega = 1$	neutral evolution (synonymous and nonsynonymous substitutions occur at the same rate)	pseudogenes
$\omega > 1$	positive selection (nucleotide substitutions often change the amino acid)	envelope proteins in viruses under active positive selection